

UNIVERSITY OF MYSORE



Estd. 1916

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No.AC.2(S)/384/14-15

Dated: 10-06-2015

NOTIFICATION

Sub: Changes in the Scheme of study and paper from the Soft core to Hard core from the Academic Year 2015-16.

Ref: 1. Proceedings of Faculty of Science & Technology Meeting held on 02-02-2015.
2. Proceedings of the Meeting of Academic Council held on 27-03-2015.

The Board of Studies in Biotechnology (PG) at its meeting held on 20-11-2014 has resolved to change the paper: "Immunotechnology" from soft core to hard core, and Paper: "Biophysics & structural biology" from soft core to hard core.

The Practical component of Hard Core Paper: "Plant Biotechnology" has been strengthened by addition of new practical experiments.

The Faculty of Science and Technology and the Academic Council at their meetings held on 02-02-2015 and 27-03-2015 respectively have approved the above proposals and the same is notified.

The copy of Changes in the Scheme of study and paper from the Soft core to Hard core is annexed.

DRAFT APPROVED BY THE REGISTRAR

[Handwritten signature]
REGISTRAR
16/6
12/6/15
4
12/6

To

1. The Registrar (Evaluation), University of Mysore, Mysore.
2. The Chairperson, BOS/DOS in Biotechnology, MGM.
3. The Dean, Faculty of Science & Technology, DOS in Earth Science, MGM.
4. The Principals of the Affiliated Colleges running MSc in Biotechnology.
5. The Director, College Development Council, UOM, Mysore.
6. The Coordinator, Online & Outreach programme, Parakalamatta, MGM.
7. The Deputy/Assistant Registrar (Evaluation), University of Mysore, Mysore.
8. The Supdt., A.B., Academic Section /PMEB, UOM., Mysore.
9. The P.A. to the Vice-Chancellor/Registrar/Registrar(Evaluation), UOM., Mysore.
10. The Case Worker, AC.7, Academic Section, University of Mysore, Mysore.
11. The Section Guard File(Supdt.AC.2), A.B., A.C., UOM.
12. The Schedule File.

University of Mysore
Department of Studies in Biotechnology
Manasagangotri, Mysore – 570 006

Scheme of Study – Revised (Academic Year 2015-16)

Master's Degree Program in Biotechnology

| | |
|-----------------------------|------------|
| Credits to be earned | 76 |
| Core papers | 52 credits |
| Soft core | 20 credits |
| Open elective paper* | 04 credits |

*Open elective shall be entirely from different discipline of study

Credit matrix for Master's Degree Program in Biotechnology

| Credits to be earned | I | II | III | IV | Total Credits |
|-----------------------------|-----------|-----------|------------|-----------|----------------------|
| Hard Core | 12 | 12 | 18 | 10 | 52 credits |
| Soft Core | 04 | 04 | 04 | 08 | 20 credits |
| Open elective | - | 04 | - | - | 04 credits |
| Total | 16 | 20 | 22 | 18 | 76 |

I Semester

| Paper Code | Title of the course | HC/SC/OE/E/Pr./etc | L | T | P | Credits |
|--|-----------------------------------|---------------------------|----------|----------|----------|----------------|
| | Bioanalytical Techniques | HC | 3 | 1 | 0 | 4 |
| | Microbiology | HC | 3 | 1 | 0 | 4 |
| | Practical-1 | HC | 0 | 0 | 4 | 4 |
| SOFTCORE (Choose any ONE from the below listed) | | | | | | |
| | Enzymology & Metabolism | SC | 3 | 1 | 0 | 4 |
| | Biophysics and Structural Biology | SC | 3 | 1 | 0 | 4 |

II Semester

| Paper Code | Title of the course | HC/SC/OE/E/Pr./etc | L | T | P | Credits |
|-------------------|----------------------------|---------------------------|----------|----------|----------|----------------|
| | Advanced Molecular Biology | HC | 3 | 1 | 0 | 4 |
| | Gene Technology | HC | 3 | 1 | 0 | 4 |

| | | | | | | |
|--|---|----|---|---|---|---|
| | Practical-2 | HC | 0 | 0 | 4 | 4 |
| SOFTCORE (Choose any ONE from the below listed) | | | | | | |
| | Molecular Genetics | SC | 3 | 1 | 0 | 4 |
| | Food & Environmental Biotechnology | SC | 3 | 1 | 0 | 4 |
| | OPEN ELECTIVE (Choose from other department) | OE | | | | 4 |
| | Applied Biotechnology (For other discipline students) | OE | 3 | 1 | 0 | 4 |

III Semester

| Paper Code | Title of the course | HC/SC/OE /E/ Pr./etc | L | T | P | Credits |
|--|--------------------------------|----------------------|---|---|---|---------|
| | Plant Biotechnology | HC | 3 | 1 | 0 | 4 |
| | Animal Biotechnology | HC | 3 | 1 | 0 | 4 |
| | Immunotechnology | HC | 3 | 1 | 0 | 4 |
| | Practical-3 | HC | 0 | 0 | 6 | 6 |
| SOFTCORE (Choose any ONE from the below listed) | | | | | | |
| | Biostatistics & Bioinformatics | SC | 3 | 1 | 0 | 4 |
| | Seed Health & Diagnostics | SC | 3 | 1 | 0 | 4 |

IV Semester

| Paper Code | Title of the course | HC/SC/OE /E/ Pr./etc | L | T | P | Credits |
|--|---------------------------------|----------------------|---|---|---|---------|
| SOFTCORE (Choose any TWO from the below listed) | | | | | | |
| | Bioprocess Technology | SC | 3 | 1 | 0 | 4 |
| | Cell Signalling & Communication | SC | 3 | 1 | 0 | 4 |
| | Cancer Biology | SC | 3 | 1 | 0 | 4 |
| | Molecular Phytobacteriology | SC | 3 | 1 | 0 | 4 |
| | Dissertation* | HC | 0 | 4 | 6 | 10 |

*** Dissertation should be in-house only and should be allotted to the students in the III Semester itself.**

Note:

Each course (Theory) = 48 h per Semester (3h per weeks)
 Each course (Practical) = 192 h per Semester (12h per week)
 Each course (Tutorial) = 32 h per Semester (2h per week)
 Dissertation = 320 h per Semester (20h per week)

M.Sc Biotechnology
Semester III
Practical -3 (HC)

| Existing | Modified |
|---|---|
| <p style="text-align: center;">Plant Biotechnology</p> <ul style="list-style-type: none"> • Preparation of plant tissue culture media • Organ cultures: Shoot tip, nodal, anther and leaf cultures • Protoplast isolation technique • Synthetic seeds • TLC of plant secondary metabolites • Alkaloid estimation • Seed structure • <i>Agrobacterium</i> culture, transformation and selection of transformants • GUS expression in transformed tissues <p style="text-align: center;">Animal Biotechnology</p> <ul style="list-style-type: none"> • Preparation of media, culture and maintenance of cell lines, trypsinization • Culture of transformed cells • MTT assay for cytotoxicity • ³H-Thymidine uptake assay for cell proliferation • Cryopreservation and revival of cells • Transient transfection assay using RSV gal gene for transfer • In vitro growth of blood vessels • Lymphocyte preparation <p style="text-align: center;">Immunotechnology</p> <ul style="list-style-type: none"> • Preparation of antigen and antibody production. • Purification of IgG. • Slide agglutination test/ Blood grouping. • Immunoprecipitation test- Ouchterlony double diffusion. • Immunoaffinity purification of IgG. • Immunofluorescence for localization of an antigen. • ELISA for quantification of an antigen. • Rossette assay. • Assay for activation of phagocytic cells. • Western blotting and detection. | <p style="text-align: center;">Plant Biotechnology</p> <ul style="list-style-type: none"> • Preparation of plant tissue culture media • Callus induction • Induction of somatic embryogenesis • Establishment of cell suspension cultures for plant secondary metabolite production • Encapsulation of somatic embryos and production of synthetic seeds • Organ cultures: Shoot tip, nodal, anther and leaf cultures • Micropropagation technique • Protoplast isolation technique • Secondary metabolite estimations: Colorimetry and TLC methods • Seed Structure • <i>Agrobacterium</i>-mediated genetic transformation • GUS expression in transformed tissues <p style="text-align: center;">Animal Biotechnology</p> <ul style="list-style-type: none"> • Preparation of media, culture and maintenance of cell lines, trypsinization • Culture of transformed cells • MTT assay for cytotoxicity • ³H-Thymidine uptake assay for cell proliferation • Cryopreservation and revival of cells • Transient transfection assay using RSV gal gene for transfer • In vitro growth of blood vessels • Lymphocyte preparation <p style="text-align: center;">Immunotechnology</p> <ul style="list-style-type: none"> • Preparation of antigen and antibody production. • Purification of IgG. • Slide agglutination test/ Blood grouping. • Immunoprecipitation test- Ouchterlony double diffusion. • Immunoaffinity purification of IgG. • Immunofluorescence for localization of an antigen. • ELISA for quantification of an antigen. • Rossette assay. • Assay for activation of phagocytic cells. • Western blotting and detection. |

